

AD_____

AWARD NUMBER: W81XWH-05-1-0592

TITLE: PSMA-Targeted Nano-Conjugates as Dual-Modality (MRI/PET) Imaging
Probes for the Non-Invasive Detection of Prostate Cancer

PRINCIPAL INVESTIGATOR: Xiankai Sun, Ph.D.

CONTRACTING ORGANIZATION: University of Texas Southwestern Medical Center
at Dallas
Dallas, TX 75390

REPORT DATE: January 2010

TYPE OF REPORT: Final Addendum

PREPARED FOR: U.S. Army Medical Research and Materiel Command
Fort Detrick, Maryland 21702-5012

DISTRIBUTION STATEMENT: Approved for Public Release;
Distribution Unlimited

The views, opinions and/or findings contained in this report are those of the author(s) and should not be construed as an official Department of the Army position, policy or decision unless so designated by other documentation.

REPORT DOCUMENTATION PAGE				<i>Form Approved</i> OMB No. 0704-0188	
<small>Public reporting burden for this collection of information is estimated to average 1 hour per response, including the time for reviewing instructions, searching existing data sources, gathering and maintaining the data needed, and completing and reviewing this collection of information. Send comments regarding this burden estimate or any other aspect of this collection of information, including suggestions for reducing this burden to Department of Defense, Washington Headquarters Services, Directorate for Information Operations and Reports (0704-0188), 1215 Jefferson Davis Highway, Suite 1204, Arlington, VA 22202-4302. Respondents should be aware that notwithstanding any other provision of law, no person shall be subject to any penalty for failing to comply with a collection of information if it does not display a currently valid OMB control number. PLEASE DO NOT RETURN YOUR FORM TO THE ABOVE ADDRESS.</small>					
1. REPORT DATE 1 January 2010		2. REPORT TYPE Final Addendum		3. DATES COVERED 15 Sep 2009 – 14 Dec 2009	
4. TITLE AND SUBTITLE PSMA-Targeted Nano-Conjugates as Dual-Modality (MRI/PET) Imaging Probes for the Non-Invasive Detection of Prostate Cancer				5a. CONTRACT NUMBER	
				5b. GRANT NUMBER W81XWH-05-1-0592	
				5c. PROGRAM ELEMENT NUMBER	
6. AUTHOR(S) Xiankai Sun, Ph.D. E-Mail: Xiankai.Sun@UTSouthwestern.edu				5d. PROJECT NUMBER	
				5e. TASK NUMBER	
				5f. WORK UNIT NUMBER	
7. PERFORMING ORGANIZATION NAME(S) AND ADDRESS(ES) University of Texas Southwestern Medical Center at Dallas Dallas, TX 75390				8. PERFORMING ORGANIZATION REPORT NUMBER	
9. SPONSORING / MONITORING AGENCY NAME(S) AND ADDRESS(ES) U.S. Army Medical Research and Materiel Command Fort Detrick, Maryland 21702-5012				10. SPONSOR/MONITOR'S ACRONYM(S)	
				11. SPONSOR/MONITOR'S REPORT NUMBER(S)	
12. DISTRIBUTION / AVAILABILITY STATEMENT Approved for Public Release; Distribution Unlimited					
13. SUPPLEMENTARY NOTES					
14. ABSTRACT The goal of this project is to develop dual modality imaging probes for the detection of prostate cancer by doping radioisotopes to iron oxide nanoparticles, so that the sensitivity and specificity of prostate cancer diagnosis could be significantly improved. During the funding period, a facile approach was developed to prepare gamma- or positron emitting nuclides incorporated SPIO nanoparticles (NUSPIONS) for dual modality imaging of prostate cancer. In this three-month extension, we successfully developed the logistics of importing ⁷⁴ As/GeO ₂ targets from Europe and developed a standardized operation protocol for the isotope separation and ⁷⁴ AsI ₃ preparation. Using ⁷⁴ AsI ₃ , we have applied our developed methodology to synthesize ⁷⁴ As incorporated dextran-coated iron oxide nanoparticles (~ 20 nm) and performed PET-CT and MRI evaluation in a PC-3 tumor mouse model. The PET/MRI dual modality imaging results clearly demonstrated the practicality of the methodology conceived and developed in this project for non-invasive detection of prostate cancer. Further research is warranted to move this project forward.					
15. SUBJECT TERMS None provided.					
16. SECURITY CLASSIFICATION OF:			17. LIMITATION OF ABSTRACT UU	18. NUMBER OF PAGES 8	19a. NAME OF RESPONSIBLE PERSON USAMRMC
a. REPORT U	b. ABSTRACT U	c. THIS PAGE U			19b. TELEPHONE NUMBER (include area code)

Table of Contents

	<u>Page</u>
Introduction.....	3
Body.....	3
Key Research Accomplishments.....	6
Reportable Outcomes.....	7
Conclusion.....	7

Introduction

As mentioned in my letter of request for this three-month extension, we ordered two shipments of As-74 from Belgium for this project. In the Revised SOW and Timeline, we specified one task to be performed in this three-month period:

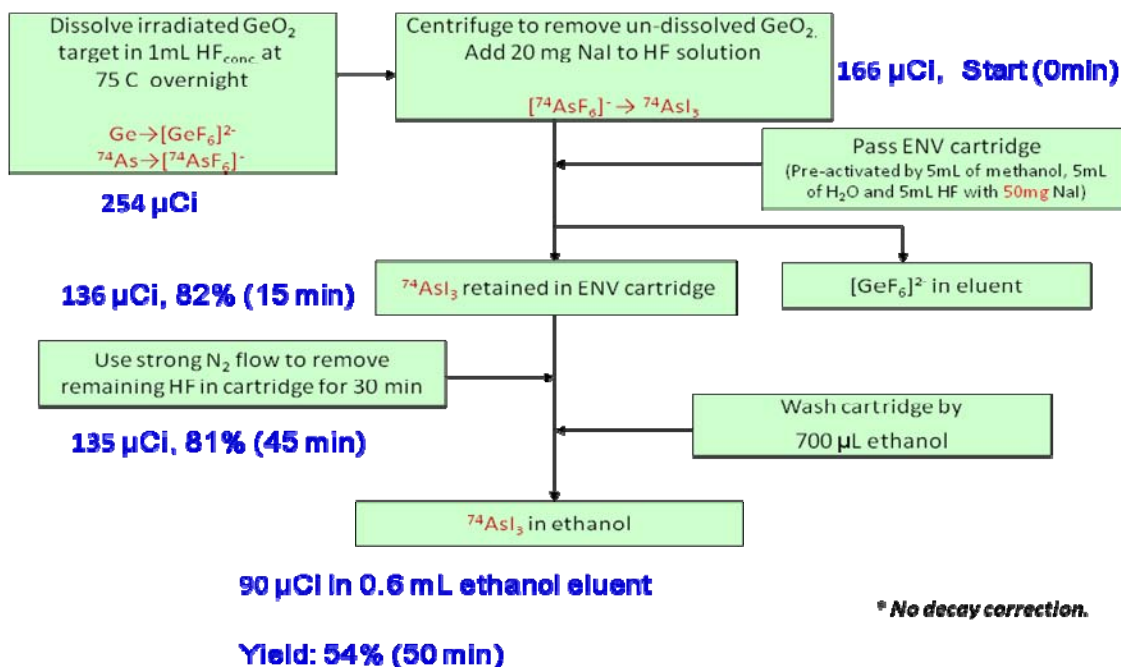
Months 49 – 51 (Task 2/Subtask 2):

We have requested two shipments of ^{74}As from Belgium to “Accomplish the small animal MRI and PET imaging evaluation of the two nano-conjugates based on ^{74}As -incorporated iron oxide nanoplatform.”

Research Progress in the three-month extension:

1. Target Processing of Irradiated $^{74}\text{As}/\text{GeO}_2$ powder for $^{74}\text{AsI}_3$ Preparation

Two shipments of $^{74}\text{As}/\text{GeO}_2$ were received in the three-month extension from the VUB-Eenheid Cyclotron Facility of Vrije Universiteit Brussels, Belgium. The activity of ^{74}As was 3.93 mCi for the first shipment (11/30/09) and 4.3 mCi for the second shipment (12/7/09). The ^{74}As activity was in 350 mg of irradiated GeO_2 powder. After several trials, we have established a standardized operation procedure for the separation of ^{74}As from GeO_2 and preparation of $^{74}\text{AsI}_3$ in reasonable radioactivity recovery rates as shown in **Scheme 1**.



Scheme 1. Schematic $^{74}\text{As}/\text{GeO}_2$ target processing and preparation of $^{74}\text{AsI}_3$

2. Incorporation of ^{77}As into dextran coated iron oxide nanoparticles

The synthetic procedure was adapted from the standard protocol of incorporating ^{77}As into iron oxide nanoparticles (see the 1st annual report) as follows.

The Fe_3O_4 nanoparticles incorporated with $^{74}\text{As}^{3+}$ were obtained via co-precipitation of Fe^{2+} , Fe^{3+} , and radioisotope in a solution phase system by the addition of a base solution. First, 35.7mg of $\text{FeCl}_3 \cdot 6\text{H}_2\text{O}$, 13.1mg of $\text{FeCl}_2 \cdot 4\text{H}_2\text{O}$, and 2.0g of Dextran T10 were dissolved in 30 mL deoxygenated Milli-Q water by intensive stirring under a protection of N_2 gas flow until a homogenous transparent light yellow solution with a pH value of 3 was obtained. The following reactions were carried out in a three-neck flask under the protection of a N_2 gas with flow rate at 2 LPM. To 20 ml of the $\text{Fe}^{3+}/\text{Fe}^{2+}$ /dextran solution, 70.4 μCi of $^{74}\text{AsI}_3$ in 1.0 mL ethanol was added. The flask was immersed in an ice/water box and kept stirring at 800 rpm for 15 min. Then 1 mL of 6.55 wt.% ammonium hydroxide solution was added slowly and continuously at a rate of 6mL/h by the control of a syringe pump. The light yellow solution slowly turned into dark green then to black. After stirring the solution for 30 min, the flask was transferred from the ice/water box to an oil bath and subsequently heated up to 78°C in an hour. A pH value of 7 was measured after the solution cooled down to room temperature. The cross linking of dextran shell was carried out by adding 6 mL of 5 M NaOH and 3mL of epichlorohydrin. The reaction was allowed to proceed for 2 h at room temperature. The resulted reaction mixture with total radioactivity of 68.4 μCi was concentrated by centricon filter tubes to remove the non-reacted ions and excess dextran molecules. The nanoparticles (DLS diameter: 20 nm) were washed several times with water until less than 1 μCi of radioactivity was detected from the filtrate. The yield of ^{74}As incorporation was 30.5% after decay correction. The injection doses were prepared in 400 μL of solution with 11.7 μCi of ^{74}As -activity and 0.1 mmol/kg iron.

3. PET-CT and MRI Imaging of Prostate Cancer with ^{74}As -incorporated Iron Oxide Nanoparticles

Tissue Culture and Animal Model: All animal studies were performed in compliance with guidelines set by the UT Southwestern Institutional Animal Care and Use Committee. The PC-3 cell line was obtained from the American Type Culture Collection (ATCC, Manassas, VA). PC-3 cells were cultured in T-media (Invitrogen, Carlsbad, CA) at 37°C in an atmosphere of 5% CO_2 and were passaged at 75 % confluence in P150 plates. T-media was supplemented with 5% Fetal Bovine Serum (FBS) and 1 \times Penicillin/Streptomycin. PC-3 cells were harvested from monolayer using PBS and trypsin/EDTA, and suspended in T-media with 5% FBS. The cell suspension was then mixed 1:1 with MatrigelTM and injected subcutaneously (2.5×10^6 cells per injection, injection volume 100 μL) into both front flanks of

SCID mice. After injection, animals were monitored three times a week by general observations. The tumor was noticed to grow in the first week and allowed to grow three weeks to reach a palpable size for the imaging experiments.

MicroPET-CT: Small animal PET-CT imaging studies were performed on a Siemens Inveon PET-CT Multimodality System (Siemens Medical Solutions Inc., Knoxville, TN, USA) when the prostate cancer xenografts reached the similar size of approximate 100 mm^3 . The injected dose was $11.7 \text{ } \mu\text{Ci}$ of ^{74}As -activity in $400 \text{ } \mu\text{L}$ PBS (three injections); and the iron dose was kept at 0.1 mmol/kg for MRI evaluation.

Ten minutes prior to imaging, the animal was anesthetized using 3% Isoflurane at room temperature until stable vitals were established. Once the animal was sedated, the animal was placed onto the imaging bed under 2% Isoflurane anesthesia for the duration of the imaging. The microCT imaging was acquired at 80 kV and 500 A with a focal spot of $58 \text{ } \mu\text{m}$. The total rotation of the gantry was 360° with 360 rotation steps obtained at an exposure time of approximately 235 ms/frame . The images were attained using a CCD readout of 4096×3098 with a bin factor of 4 and an average frame of 1. Under low magnification the effective pixel size was $103.03 \text{ } \mu\text{m}$. Total microCT scan time was

approximately 6 min. CT images were reconstructed with a down sample factor of 2 using Cobra Reconstruction Software. The PET imaging was acquired directly following the acquisition of CT data. The PET tracers were injected intravenously via the tail vein. Static PET scans were performed at 15 h post injection for 45 min. PET images were reconstructed using Fourier Rebinning and Ordered Subsets Expectation Maximization 3D (OSEM3D) algorithm. Reconstructed CT and PET images were fused and analyzed using the Siemens Inveon Research Workplace (IRW) software.

The PET-CT images of a mouse injected with ^{74}As -incorporated iron oxide nanoparticles are shown in Figure 1. The PC-3 tumor showed

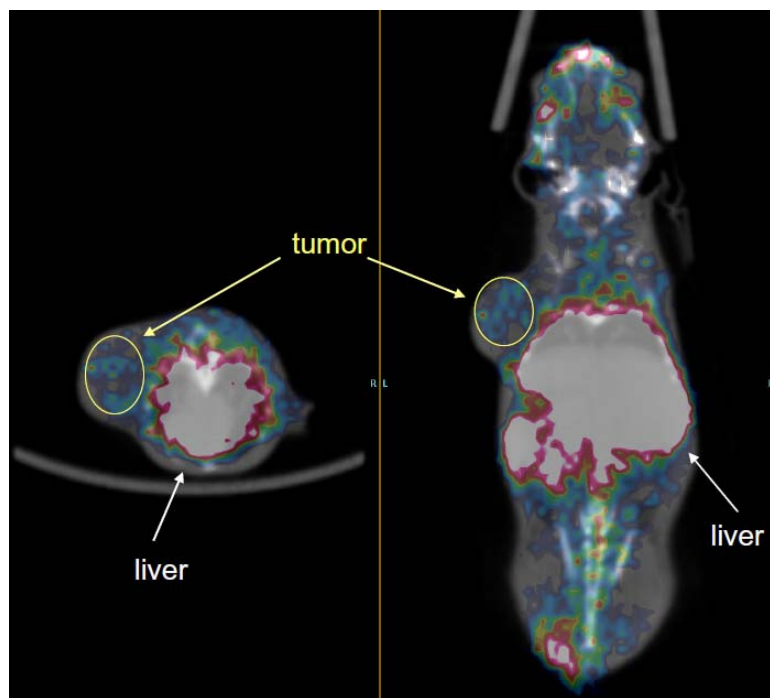


Figure 1. PET-CT images of a PC-tumor bearing mouse injected with ^{74}As -incorporated iron oxide nanoparticles. Left: transaxial; right: coronal. The imaging was acquired for 45-min after 15 h post-injection.

reasonable uptake of the nanoparticles at 12-h post-injection. Due to the low injection of ^{74}As -activity, the imaging was acquired for 45-min. The tumor PET contrast (signal/noise) may be improved by increasing the injection dose of ^{74}As -activity. The high liver accumulation reflects the nature of nanoparticles in vivo, indicating the necessity of nanoparticle surface modifications.

Mouse MRI. The MRI evaluation was performed on a 4.7 T Varian MRI system equipped with a homemade coil after the completion of PET-CT imaging. The mice were anesthetized by inhalation of 1.5-2.0 % isoflurane in air. Multi-slice T_2 maps were obtained using a spin echo sequence.

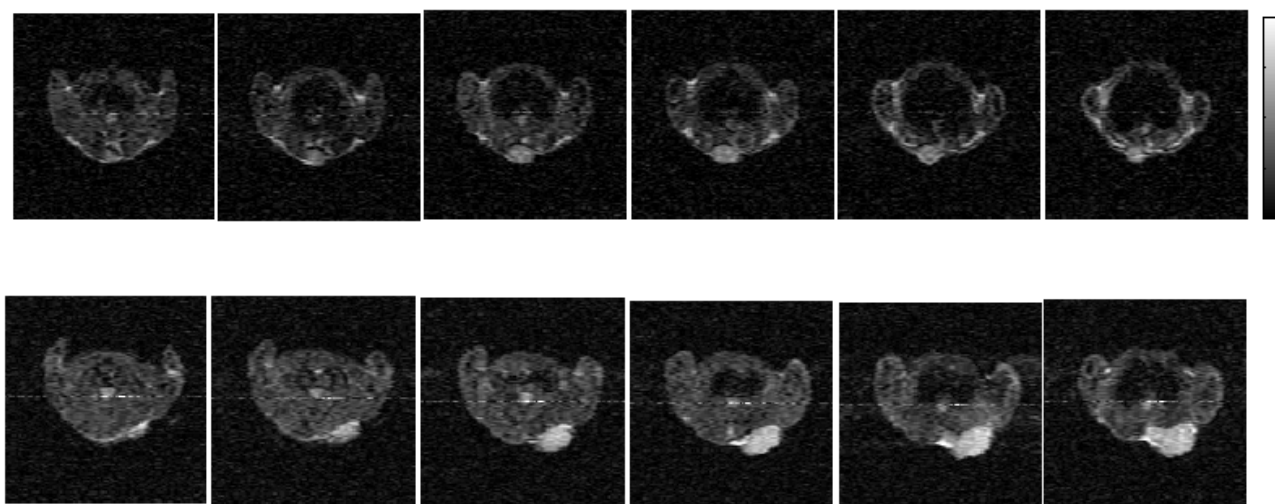


Figure 2. Multi-slice T_2 MR images of a PC-3 tumor-bearing mouse injected with ^{74}As -incorporated iron oxide nanoarticles at the iron dose of 0.1 mmol/kg (top panel) and a PC-3 tumor-bearing mouse without the nanoparticle injection (low panel).

As shown in Figure 2, we can clearly see the T_2 shortening effect resulted from the tumor (top) with iron oxide nanoparticle accumulation as compared to the control tumor (bottom). Therefore we performed ROI analysis on a slice-by-slice basis using homebuilt Matlab routines and the T_2 values from all voxels of each tumor were pooled for statistical comparison using either students' t-test or 1-way ANOVA by GraphPad Prism® program. We observed statistically significant decrease (compared to Control tumor) in the mean T_2 value from 52.7 ± 14.0 ms to 43.6 ± 12.5 ms in mice injected with iron oxide nanoparticles.

Key Research Accomplishments

In addition to the accomplishments summarized in my annual and final reports, we have achieved the following ones in this three-month extension:

- 1) Successfully separated ^{74}As from $^{74}\text{As}/\text{GeO}_2$ targets and established a standard procedure to synthesize $^{74}\text{AsI}_3$.
- 2) Successfully applied our developed methodology to incorporate ^{74}As into the core of dextran-coated iron oxide nanoparticles (~ 20 nm).
- 3) Successfully performed PET-CT and MRI evaluation of the ^{74}As -incorporated iron oxide nanoparticles in a PC-3 tumor mouse model. PET imaging of PC-3 tumor with ^{74}As -incorporated iron oxide nanoparticles showed a marginal success, while T_2 weighted MRI results were much more significant.

Reportable Outcomes

One manuscript will be prepared based on the results obtained in this extended period.

Conclusions

In this three-month extension, we have accomplished the preparation of ^{74}As -incorporated iron oxide nanoparticles and successfully used the PET/MRI dual modality imaging probes to visualize prostate cancer in a subcutaneous tumor mouse model. While this part of work was impeded by the availability of ^{74}As , to date we have shown the practicality of using ^{74}As -incorporated iron oxide nanoparticles for PET/MRI dual modality imaging of prostate cancer. Therefore I believe we have satisfactorily accomplished the goal set for this project.